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DOI:

[10.1177/0333102418756863](https://doi.org/10.1177/0333102418756863)

*Document Version*

Peer reviewed version

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*Citation for published version (APA):*

Holland, P. R., Saengjaroentharn, C., & Vila-Pueyo, M. (2018). The role of the brainstem in migraine: Potential brainstem effects of CGRP and CGRP receptor activation in animal models. *Cephalalgia*.  
<https://doi.org/10.1177/0333102418756863>

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# The role of the brainstem in migraine: Potential brainstem effects of CGRP and CGRP receptor activation in animal models

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## **Abstract**

**Background:** Migraine is a severe debilitating disorder of the brain that is ranked as the 6<sup>th</sup> most disabling disorder globally, with respect to disability adjusted life years and there remains a significant unmet demand for an improved understanding of its underlying mechanisms. In conjunction with perturbed sensory processing, migraine sufferers often present with diverse neurological manifestations (premonitory symptoms) that highlight potential brainstem involvement. Thus, as the field moves away from the view of migraine as a consequence of purely vasodilation to a greater understanding of migraine as a complex brain disorder, it is critical to consider the underlying physiology and pharmacology of key neural networks likely involved.

**Discussion:** The current review will therefore focus on the available evidence for the brainstem as a key regulator of migraine biology and associated symptoms. We will further discuss the potential role of CGRP in the brainstem and its modulation for migraine therapy, given the emergence of targeted CGRP small molecule and monoclonal antibody therapies.

**Conclusion:** The brainstem forms a functional unit with several hypothalamic nuclei that are capable of modulating diverse functions including migraine-relevant trigeminal pain processing, appetite and arousal regulatory networks. As such, the brainstem has emerged as a key regulator of migraine and is appropriately considered as a potential therapeutic target. While currently available CGRP targeted therapies have limited blood brain barrier penetrability, the expression of CGRP and its receptors in several key brainstem nuclei and the demonstration of brainstem effects of CGRP modulation highlight the significant potential for the development of CNS penetrant molecules.

**Keywords:** Migraine, Brainstem, CGRP.

## **Introduction**

Migraine is a severe debilitating disorder of the brain that has troubled humans for centuries (1). Despite representing the 6<sup>th</sup> most disabling disorder globally (2), and the most disabling neurological condition with respect to disability adjusted life years, there remains a significant unmet demand for an improved understanding of the underlying disease mechanisms. Several theories have been proposed for migraine-related pain, including vasodilation (3, 4), neurogenic inflammation (5) and activation of the trigeminal pain processing primary afferent nociceptors that innervate the intra- and extra-cranial structures of the head (3, 6). In conjunction with the developing clinical picture, whereby migraine is considered a multiphasic disorder, current theories of migraine propose a complex neural origin with subsequent vascular involvement (6, 7).

While head pain is a seminal characteristic of migraine (8) patients often present with several associated neurological symptoms. These include perturbed sensory responses to light (photophobia), sound (phonophobia), smell (osmophobia) and touch (allodynia) (9). In addition to this abnormal sensory processing, cognitive, emotional and motor abnormalities are commonly reported by migraine patients that may be confounded by several comorbid conditions including insomnia (10) and depression (11). Given the diversity of symptoms, migraine likely involves the interaction of several neural networks that regulate these diverse functions (e.g. nociception, arousal and appetite) (7). As evidenced by the presence of several common premonitory (prodromal) symptoms (12) that include disrupted homeostatic mechanisms regulating sleep/wake and appetite regulation (e.g. fatigue, yawning, loss of appetite). Thus, as the migraine field moves away from the view of migraine as a consequence purely of vasodilation (13-15) to a greater understanding of migraine as a brain disorder (1, 6, 7), it is critical to consider the underlying pathways known to be involved in the diverse symptoms of migraine. In doing so, it is further

important to consider if anti-migraine therapeutic agents, such as targeted calcitonin gene-related peptide (CGRP) modulation, may in part act via these networks? As such, the current review will focus on the available evidence for the brainstem as a key regulator of migraine biology. We will further discuss the potential role of CGRP in the brainstem and its modulation for migraine therapy, given the emergence of targeted CGRP small molecule and monoclonal antibody therapies.

### **Migraine-related pain processing and the brainstem**

Undoubtedly, one of the most salient features of migraine is the severe debilitating often unilateral pulsating head pain that is characteristic of the condition. This is borne from the activation, or perceived activation, of the pain processing trigeminovascular system that processes sensory information from the intra- and extra-cranial structures of the head and face (Figure 1). The exact role of trigeminovascular activation is still debated. Does an underlying CNS disruption result in the diverse symptoms associated with migraine, whilst driving aberrant trigeminovascular activation, or is trigeminovascular activation sufficient to trigger an attack and drive the diverse symptoms? Irrespective of the sequence of events, it is clear that the pain processing trigeminovascular system and its central projections represent a key interface between migraine-related pain and its associated non-pain features (Figure 2). As such, activation of the trigeminovascular system has been consistently used as a preclinical model of migraine-related pain processing (nociception). Primary sensory afferents arising in the trigeminal ganglion (TG) project peripherally to the intra- and extracranial structures, including the pial and dural blood vessels (3, 16, 17), as well as projecting centrally to the trigeminal nucleus caudalis and its cervical boundaries (18-23), giving rise to the trigeminocervical complex (TCC) (Figure 1). From the TCC, second order neurons ascend in the trigemino- and quintothalamic tracts where they primarily

synapse on ventral posteromedial nucleus of the thalamus (24-28), before being widely distributed throughout the cortex for the integration of sensory, affective and cognitive aspects of pain. Critically, these ascending trigeminothalamic projections form additional connections with areas of the brainstem including the periaqueductal grey (PAG), rostral ventromedial medulla (RVM), nucleus raphe magnus (MRN) and the locus coeruleus (LC) (24, 29), as well as to various hypothalamic nuclei (30-34).

The importance of the trigeminal-brainstem interactions is highlighted in several neuroimaging studies that demonstrate brainstem activation during spontaneous (35) and triggered attacks (36). Whilst this activation was originally suggested to be in response to the developing headache, recent observations have highlighted that brainstem activation occurs during the earliest premonitory phases (37) prior to the headache. This is further supported by the demonstration of altered brainstem functional connectivity (hypothalamus-PAG) in a temporally defined phase prior to the onset of spontaneous migraine attacks (38). Despite the clear evidence for brainstem involvement, the specific brainstem structures involved are still debated due to the limited resolution of neuroimaging approaches. Further, it is not clear if this altered brainstem activation is occurring independent of an increasing activation of the trigeminovascular system, as it has been shown that the TCC increases its responsiveness to noxious intranasal stimuli as the next migraine attack approaches (39). The brainstem may act as a component of a homeostatic regulatory network that monitors and reacts to increasing trigeminovascular activity in response to endogenous and exogenous signals that can predispose an individual to attacks.

In addition to receiving direct ascending TCC projections and being activated in response to nociceptive durovascular stimulation in animal models (19, 40-44), these brainstem structures, that likely include the PAG, LC, RVM and MRN are intrinsically linked throughout the

trigeminovascular system, including to higher order structures such as the hypothalamus and thalamus (Figure 1) (1). This diverse network of brainstem nuclei is ideally positioned to integrate trigeminal mediated signals with neural networks regulating migraine-associated symptoms (Figure 2) and represents a potential target for neuromodulatory and pharmacological approaches for migraine.

### **The Brainstem and homeostatic regulation**

The hypothalamus has emerged as a key regulator of homeostatic mechanisms (45), sensing intrinsic and extrinsic signals while orchestrating appropriate behavioural responses. While the hypothalamus resides within the diencephalon, it is intrinsically linked with several brainstem nuclei forming a functional homeostatic regulatory network. As such, the brainstem plays a significant role in multiple migraine-relevant biological functions (as detailed in Figure 2). For example, they play a key role in the integration of local and circulating factors that convey information regarding energy balance acting to regulate appetite and energy expenditure (46). Vagal afferents from the gastrointestinal tract signal via the nucleus of the solitary tract (NTS) and the area postrema (AP; also plays a role in nausea (47)) to the hypothalamic nuclei regulating appetite. Critically, this appetite-regulating pathway, including the AP and NTS has been shown to express functional CGRP receptor components (Figure 3 and Table 1), while lesions in the area of the AP/NTS abolishes CGRP induced anorectic effects (48). It is noteworthy that the AP is devoid of the BBB (49), suggesting it is amenable to peripherally administered CGRP therapies; however, the functional relevance of this localized BBB permeability remains to be determined (see section on CGRP and the BBB).

Further, several brainstem nuclei regulate arousal levels (Figure 2) and therefore have been linked to the presence of abnormal fatigue-like symptoms during migraine (12, 50). The majority of these structures, including the PAG and LC, receive wake-promoting projections from the hypothalamus (for review see (51)), contain CGRP fibers/cell bodies and express CGRP receptor components (52-55) (see Figure 3 and Table 1 for detailed distribution). Given their established roles in the modulation of trigeminovascular activity and cerebral blood flow (56-59) they likely form a key interface between migraine and its association with sleep/wake disturbances (60). As such, the brainstem is ideally positioned to regulate several biological functions related to migraine, including appetite and fatigue (arousal dysregulation), is abnormally active during the earliest attack phases and has been proposed to play a role in attack initiation.

**Table 1. CGRP receptor expression, CGRP binding patterns and CGRP expression in selected brainstem nuclei.**

The data is further presented in figure form in figure 3. AP; area postrema, CB; cell bodies, CLR; calcitonin receptor-like receptor, DMV; dorsal motor nucleus of the vagal, DR; dorsal raphe, F; Fibers, LC; locus coeruleus, MK-3207; CGRP receptor antagonist, Mod; moderate, MRN, nucleus raphe magnus, NTS; nucleus of the solitary tract, PAG; periaqueductal gray, RAMP1: receptor activity modifying protein 1, RCP, receptor component protein, SuS; superior salivatory nucleus, TCC; trigeminocervical complex, VN; vestibular nuclei



	<b>CGRP receptors components/CGRP Binding</b>	<b>CGRP Expression</b>	<b>Species</b>	<b>Ref</b>
AP	CLR & RAMP1 (Protein & mRNA); MK-3207 Binding N/A	N/A F (High)	Primate Alpaca	(53) (61)
DMV	RAMP1 (F) RCP (Mod CB) N/A CLR & RAMP1 (Protein & mRNA); MK-3207 Binding GRP Binding (High)	CB CB (Mod) F (Low); CB (High) N/A N/A	Rat Rat Alpaca Primate Human/Rat	(62) (54) (61) (53) (63)
DR	RAMP1 (mRNA); CLR & RAMP1 (Protein); MK-3207 Binding CGRP Binding (Mod) N/A RCP (Mod Cell Bodies)	N/A  N/A F (Low); CB (High) Axons & CB (Low)	Primate  Human Alpaca Rat	(53)  (63) (61) (54)
LC	N/A RAMP1 & CLR (F & CB) N/A RCP (Mod CB) CGRP Binding (High)	CB (80%); F (Low) CB F (Mod) CB (Mod) N/A	Human Rat Alpaca Rat Human/Rat	(55) (62) (61) (54) (63)
MRN	RAMP1 & CLR (F & CB)	CB	Rat	(62)
NTS	Absent N/A RCP (Mod CB) CGRP Binding (High)	F & CB F (High) Axons (Mod) N/A	Rat Alpaca Rat Human/Rat	(62) (61) (54) (63)
PAG	CLR & RAMP1 (Protein & mRNA); MK-3207 Binding RCP (Mod CB) CLR & RAMP1 (Cell Bodies) CLR & RAMP1 mRNA (High); RCP mRNA (Mod) N/A CGRP Binding (Moderate) CGRP Binding (High)	N/A Axons (Low) N/A <i>CALCA</i> mRNA (Low) F & CB (Low) N/A N/A	Primate Rat Rat Rat Alpaca Rat Human/Rat	(53) (54) (64) (65) (61) (66) (63)
SuS	N/A	Fibers	Rat	(67)
TCC	CLR & RAMP1 (Protein & mRNA); MK-3207 Binding RCP (Mod CB) CLR/RAMP1 CLR mRNA (High); RAMP1 & RCP mRNA (Mod) CGRP Binding (High) CLR & RAMP1 (CB) CGRP Binding (Mod)	N/A Axons (Mod) Present (not defined) <i>CALCA</i> mRNA (Mod) N/A N/A N/A	Primate Rat Rat Rat Human Rat Rat	(53) (54) (62) (65) (63) (64) (66)
VN	N/A N/A CGRP Binding (Mod) CGRP Binding (High/Mod)	F & CB CB N/A N/A	Rat Rat Rat Human/Rat	(68) (69) (66) (63)

## **Calcitonin Gene-Related Peptide**

CGRP belongs to the calcitonin family and is synthesized from two genes; the *CALCA* gene gives rise to calcitonin or  $\alpha$ CGRP and the distinct *CALCB* gene gives rise to  $\beta$ CGRP (57). The two isoforms are similar in homology and biological function, however  $\alpha$ CGRP is classically considered to predominate in the peripheral and central nervous system, whereas  $\beta$ CGRP predominates in the enteric nervous system (70), as such we will focus on  $\alpha$ CGRP (described as CGRP from this point onward). Following the synthesis of CGRP, a process that remains poorly understood, the neuropeptide is stored in large, dense-core vesicles within nerve terminals (71).

As discussed previously, CGRP is expressed in sensory afferents innervating the cranial vasculature (72) where it can act as a potent vasodilator. However, its widespread distribution throughout the trigeminal ganglion and afferents, brainstem, diencephalic and particularly the cerebellum, highlights its diverse role in neurotransmission (73). The CGRP receptor consists of three key components: (i) the calcitonin receptor-like receptor (CLR), (ii) a receptor activity modifying protein (RAMP) and (iii) an additional receptor component protein (RCP). The CLR belongs to the same “secretin-like” family of G protein-coupled receptors as those for calcitonin, vasoactive intestinal peptide and pituitary adenylate cyclase-activating peptide and when expressed independently it is unresponsive to CGRP. CLR requires a RAMP to regulate its functional specificity and membrane translocation (73). The association of CLR with RAMP1 denotes the established CGRP receptor, CLR and RAMP2 results in the adrenomedullin receptor (AM<sub>1</sub> receptor) and CLR and RAMP3 results in an additional adrenomedullin receptor (AM<sub>2</sub> receptor). While the primary receptor for CGRP is considered to be the CLR/RAMP1 complex, it is known that *in vitro* CGRP shows partial affinity for the CLR/RAMP3 AM<sub>2</sub> receptor (74) and for a recently described third potential CGRP receptor resulting from the combination of the

calcitonin receptor and RAMP1 (75). Given the widespread distribution of CLR, it is perceived that individual RAMPs are responsible for receptor tissue (76) and species specificity (77).

The above combination of receptor proteins and RAMPs constitutes a functional receptor; however, the presence of a third RCP is essential for optimal function (78). The RCP is a small hydrophilic membrane-associated protein that enhances the signal transduction of both CGRP and adrenomedullin receptors, suggesting that it may represent a novel therapeutic target for the inhibition of all identified potential CGRP receptors.

#### Targeted CGRP Therapies and the Blood-Brain Barrier (BBB)

Currently targeted CGRP therapies are focused on either acute CGRP receptor antagonism (gepants) or prophylaxis with monoclonal antibodies that bind to the CGRP peptide or its receptor (umabs). Targeted antagonism of the CGRP receptor has demonstrated significant clinical promise (79). The initial doses required for clinical efficacy raised the possibility that the small molecules may have to penetrate the BBB to exert their effects (80). This was supported by previous animal research demonstrating that olcegepant inhibited capsaicin-induced sensitization in the TCC with little impact on the TG (81). Despite this initial focus on potential central effects, telcagepant has subsequently been shown to bind to the TG of rhesus monkeys, while failing to displace central binding of the radiolabeled CGRP PET ligand MK-4232 (82), suggesting limited CNS bioavailability in humans. This proposed peripheral site of action is further supported by the demonstration of a clear clinical efficacy for umabs (83), whose large molecular weight largely preclude BBB penetration, with only 0.1% of systemic values estimated to gain access to the CNS. In view of this relative lack of BBB permeability, it has alternatively been proposed that the BBB

is compromised during migraine attacks and, as such, the precise location of action of targeted CGRP therapies and existing migraine therapies remains a consistently debated topic. It should, however, be noted that studies utilizing radiolabeled dihydroergotamine found no evidence for BBB disruption during migraine attacks (84).

Independent of the potential disruption of the BBB during migraine, it is clear that specific brain areas lack a functional BBB and, as such, these sites offer a potential conduit for larger molecules to enter the CNS and/or facilitate signaling via widespread CNS projections. Specific areas include the circumventricular organs, meningeal arteries, pineal gland, preoptic recess and endothelium of the choroid plexus. The circumventricular organs are particularly interesting (85), these include both sensory (subfornical organ (SFO), vascular organ of the lamina terminalis (OVLT) and the AP) and secretory organs (neurohypophysis, median eminence and the pineal gland). The SFO, OVLT and AP are reciprocally interconnected with each other and with several hypothalamic and brainstem nuclei that play important roles in migraine pathophysiology. The SFO receives significant afferent input from the lateral and anterior hypothalamus, as well as the paraventricular nucleus, while its efferent projections signal to the suprachiasmatic nucleus and paraventricular nucleus. The OVLT is additionally innervated by direct projections from the dorsomedial, ventromedial and preoptic hypothalamic nuclei, as well as from the PAG and LC. It sends efferent fibers to the paraventricular, supraoptic, preoptic and lateral hypothalamic nuclei, as well as to the PAG and LC. In terms of direct brainstem access, the AP forms part of the vagal complex in conjunction with the NTS and the DMV. It is reciprocally interconnected with the other circumventricular organs in addition to the NTS and RVM, and receives direct afferent projections from the paraventricular nucleus. The AP has been shown to express CGRP receptors (Figure 3 and Table 1)(52, 53) and together with other circumventricular organs is involved in the regulation

of feeding/energy homeostasis and nausea. The functional importance for these BBB openings in relation to migraine therapy remain to be explored.

In light of this limited CNS penetrability, the concept of exploring brainstem effects of current CGRP therapies appears on the surface a fruitless endeavor. Yet, as will be discussed below, there is clear evidence for potential CNS mechanisms of CGRP modulation. This is in agreement with the demonstration of brainstem effects of triptans (58), despite variable BBB penetrability that has ultimately led to the development of a novel class of CNS penetrant, centrally acting 5-HT<sub>1F</sub> receptor agonists (ditans) (86). Taking this into consideration, it is reasonable to conclude that the development of CNS penetrant CGRP targeted therapies or novel BBB transporters remains a valid target for migraine therapy.

### **CGRP expression in the brainstem**

As detailed above,  $\alpha$ CGRP (herein denoted as CGRP) predominates in the CNS (70) and its expression has been widely mapped throughout the brainstem ((52-55), which the following section is based upon, see Figure 3 and Table 1 for a detailed breakdown of expression). Given the widespread expression of CGRP and its receptors, it has been linked to a number of functions ranging from nociception (87) to vestibular regulation and motion sickness (68), all of which have been linked to migraine. CGRP expression is largely conserved across species, including human, monkeys, cats and rodent tissue, and, as such, expression data is discussed independent of species except where notable differences have been identified. Small and medium size CGRP-expressing trigeminal primary afferents arising from the TG synapse within the superficial laminae of the TCC, where they synapse on ascending trigeminothalamic projection neurons (88). Clinical and

preclinical data consistently highlight the importance of the TCC as a key transitional interface in the processing of trigeminovascular nociception, that additionally receives descending modulatory inputs from several brainstem and higher order structures (Figure 1). The highest density of CGRP immunoreactivity in the brainstem has been reported in the LC (55, 89), with moderate expression observed in the PAG. Additional expression has been observed throughout several brainstem structures, including the dorsal raphe nuclei (DR), spinal trigeminal nuclei and vestibular nuclei (VN). Given the complexity of the CGRP receptor structure, a variety of approaches have attempted to visualize CGRP receptor distribution in the brainstem. The RCP component demonstrates high expression levels in the PAG and DR nuclei of rats, with moderate expression in the ventral tegmental area and LC; while CLR is observed in the human brainstem nuclei including the NTS and AP. In support of the widespread distribution of CGRP receptors, a recent study utilizing [ $^{11}\text{C}$ ]MK-4232 as a selective marker for the CLR/RAMP1 complex demonstrated widespread moderate distribution in the human and primate brainstems, although specific brainstem nuclei were not defined. (82).

### **Brainstem effects of CGRP and CGRP receptor modulation**

CGRP is widely distributed in the brainstem, yet relatively little is known regarding its specific function. Of particular importance to migraine is the predominance of CGRP in small to medium trigeminal primary afferents that synapse in the dorsal horn of the TCC. CGRP release at these central synapses is suggested to increase nociceptive transmission. Local delivery of CGRP onto dural nociceptive-responsive TCC neurons in the cat resulted in excitation of over 40% of neurons tested. Further, CGRP antagonism with the truncated form of CGRP (CGRP<sub>8-37</sub>) or the small molecule antagonist olcegepant (BIBN4096BS) inhibited the response of the majority of neurons

to local glutamate, suggesting a post-synaptic mechanism of action (90). In agreement, *ex vivo* studies on medullary brain slices demonstrated increased CGRP release in response to potassium chloride and capsaicin that was inhibited by naratriptan. These results highlight the importance of CGRP release from central terminals in the TCC for trigeminal nociception and suggest that TCC CGRP modulation may be a potential mechanism of action for the triptans (91).

As noted, the highest expression levels of CGRP in the brainstem are found in the noradrenergic LC (55), an area involved in the regulation of arousal (92), autonomic (93), stress (94), nociceptive functions (95) and cerebral blood flow (59, 96, 97) (Figure 2). Preclinically, the LC is responsive to activation of the trigeminovascular system (98) and known clinical migraine triggers, including nitroglycerin, induce LC activation (99); while its stimulation in the cat impacts cerebral blood flow inducing hypoperfusion (59, 96, 97) reminiscent of the functional hyperemia observed during cortical spreading depression. The LC is further activated by CTR-stimulating peptide 1, suggesting that it is sensitive to circulating CGRP levels that may act to regulate energy homeostasis (100). Inhibition of CGRP receptors with olcegepant *in vitro* in organotypic brain slice cultures containing the LC increases noradrenaline release, suggesting a possible action of CGRP targeted therapies on intrinsic CGRP signaling in the LC, which could impact on migraine pathophysiology (101). While the exact functions of the LC remain to be fully elucidated, its role as a major arousal promoting network that demonstrates clear diurnal activation patterns (102), has highlighted its potential as a regulator of migraine biology. This link has been recently strengthened by the demonstration of altered trigeminovascular dural nociceptive evoked responses following lesioning of the LC (103), whereby acute lesioning or chronic ablation of the LC reduced dural-nociceptive evoked TCC responses.

The PAG is perhaps the most prominent brainstem nuclei in migraine research since Weiller et al. (35) demonstrated its activation during a spontaneous migraine attack, with subsequent imaging studies highlighting abnormal functional activation and network connectivity during the premonitory phase (37, 38). Preclinically, this is in agreement with abnormal functional connectivity states following repetitive dural inflammatory soup application (104), suggesting that abnormal PAG signaling may be an important regulator of response to recurrent attacks. Electrical or chemical activation of the ventrolateral subunit of the PAG (vlPAG) inhibits dural vasodilation evoked trigeminal nociceptive processing in the TCC (105-107), including local application of a triptan (58). It has further been demonstrated that orally administered rizatriptan, that is thought to have a degree of central anti-nociceptive effects (108), reduces nitroglycerin-induced elevation of CGRP in the PAG (109), thus highlighting the potential of targeting such brainstem nuclei in migraine therapy. While the PAG is protected by the BBB, direct administration of CGRP into the PAG has been shown to potentiate trigeminovascular nociception at the level of the TCC, and both CGRP<sub>8-37</sub> and olcegepant inhibit TCC dural-nociceptive evoked responses in the rat (64). It must be noted, however, that direct administration of CGRP into the MRN of rats demonstrated an anti-nociceptive effect on hind-paw thermal and mechanical stimuli (110). As such, there is evidence for complex pro- and anti-nociceptive effects of CGRP signaling in brainstem nuclei that may be dependent on the specific nuclei or the pain modality being tested.

The RVM is a key output relay of the PAG involved in the indirect modulation of pain signaling. It has reciprocal connections with the PAG and TCC (111-113) and is known to contain both anti-nociceptive off and pro-nociceptive on cells (113) that can regulate trigeminovascular nociception. For example, repetitive application of an inflammatory soup to the dura mater of rats results in heightened mechanosensation via an action on RVM mediated descending facilitation (41). This



pathway has further been implicated in the regulation of homeostatic mechanisms as on-cells are state dependently active during wakefulness; however, they fall silent during feeding. Off-cells show an opposite response to feeding in that they are constitutively activated during feeding, suggesting a role in the integration of energy homeostasis (114, 115). In the context of CGRP, there is little evidence for CGRP/CGRP receptor expression within the RVM (116); however, it is likely that the RVM acts downstream of other brainstem and hypothalamic nuclei including the PAG that do show responsiveness to CGRP signaling (64).

Patients with migraine often report vertigo (117), which is regulated by the vestibular nuclei in the brainstem. CGRP is expressed throughout the various vestibular nuclei (118-120) (Figure 3 and Table 1), with preclinical studies identifying increased CGRP expression in a rotational model of motion sickness (121). Rats exposed to repeated bouts of rotary stimulation had an increased number of CGRP expressing neurons throughout the vestibular nuclei, including the vestibular efferent nucleus at the level of the facial nerve genu. This is in agreement with a reduced vestibulo-ocular reflex in CGRP null mice (122) highlighting the potential role of elevated CGRP signaling as an underlying mechanism for the association between migraine and vertigo (123).

## **Conclusion**

The brainstem forms a key functional unit with key hypothalamic nuclei that is capable of modulating diverse functions including migraine-relevant trigeminal pain processing, appetite and arousal regulatory networks. While the resolution of human neuroimaging precludes the identification of specific nuclei, it does highlight the abnormal activation of a brainstem nuclear complex during the earliest premonitory phase that continues into the headache phase and remains

after triptan-induced pain relief (35). As such, the brainstem has emerged as a key regulator of migraine biology and is appropriately considered as a potential therapeutic target. CGRP and its receptors are expressed throughout the brainstem (detailed in Figure 3 and Table 1) where their modulation has been shown to impact on trigeminal mediated pain and associated migraine-related functions. Current evidence suggests that the majority of CGRP targeted therapies, including monoclonal antibodies targeting CGRP or its receptor, do not cross the BBB in any significant quantity. Despite this lack of BBB penetration (notwithstanding localized access at areas such as the AP), there is sufficient clinical and experimental data to suggest that targeting central mechanisms including via brainstem nuclei may offer additional therapeutic benefits.

### **Article Highlights**

- The brainstem is a key region involved in the regulation of homeostatic mechanisms that play a prominent role in migraine including fatigue (arousal dysfunctions) and appetite.
- Several brainstem nuclei contain CGRP and its receptor and local application of CGRP agonists and antagonists can modulate trigeminovascular nociception.
- Modulation of brainstem CGRP signaling remains a valid target for the development of brain penetrant CGRP targeted therapies.

## **Review Process**

We searched PubMed, Medline and Web of Science with the search terms “migraine”, “brainstem”, “CGRP”, “CGRP receptor”, “periaqueductal gray”, “locus coeruleus”, “rostral ventromedial medulla”, “trigeminal” and combinations therein. We reviewed bibliographies of relevant articles and identified articles through additional author searches to include mostly preclinical articles and occasional congress proceedings/abstracts published in English. The final reference list was then selected based on the authors perception of the relevance of the article to the scope of this review.

## References

- [1] Akerman S, Holland PR, Goadsby PJ. Diencephalic and brainstem mechanisms in migraine. *Nat Rev Neurosci.* 2011;12(2011):570-84.
- [2] Murray CJ, Vos T, Lozano R, Naghavi M, Flaxman AD, Michaud C, et al. Disability-adjusted life years (DALYs) for 291 diseases and injuries in 21 regions, 1990-2010: a systematic analysis for the Global Burden of Disease Study 2010. *Lancet.* 2010;380(2010):2197-223.
- [3] Ray BS, Wolff HG. Experimental studies on headache. Pain sensitive structures of the head and their significance in headache. *Archives in Surgery.* 1940;41(1940):813-56.
- [4] Willis T. The anatomy of the brain and nerves. New York: Oxford University Press, 1964.
- [5] Moskowitz MA. Neurogenic versus vascular mechanisms of sumatriptan and ergot alkaloids in migraine. *Trends Pharmacol Sci.* 1992;13(1992):307-11.
- [6] Goadsby PJ, Holland PR, Martins-Oliveira M, Hoffmann J, Schankin C, Akerman S. Pathophysiology of Migraine: A Disorder of Sensory Processing. *Physiol Rev.* 2017;97(2017):553-622.
- [7] Burstein R, Nosedá R, Borsook D. Migraine: Multiple Processes, Complex Pathophysiology. *J Neurosci.* 2015;35(2015):6619-29.
- [8] Headache Classification Committee of the International Headache S. The International Classification of Headache Disorders, 3rd edition (beta version). *Cephalalgia.* 2013;33(2013):629-808.
- [9] Headache Classification Committee of the International Headache Society. The International Classification of Headache Disorders, 3rd edition (beta version). *Cephalalgia.* 2013;33(2013):629-808.

- [10] Uhlig BL, Engstrom M, Odegard SS, Hagen KK, Sand T. Headache and insomnia in population-based epidemiological studies. *Cephalalgia*. 2014;34(2014):745-51.
- [11] Rist PM, Schurks M, Buring JE, Kurth T. Migraine, headache, and the risk of depression: Prospective cohort study. *Cephalalgia*. 2013;33(2013):1017-25.
- [12] Giffin NJ, Ruggiero L, Lipton RB, Silberstein SD, Tvedskov JF, Olesen J, et al. Premonitory symptoms in migraine: an electronic diary study. *Neurology*. 2003;60(2003):935-40.
- [13] Amin FM, Asghar MS, Hougaard A, Hansen AE, Larsen VA, de Koning PJ, et al. Magnetic resonance angiography of intracranial and extracranial arteries in patients with spontaneous migraine without aura: a cross-sectional study. *Lancet Neurol*. 2013;12(2013):454-61.
- [14] Charles A. Migraine is not primarily a vascular disorder. *Cephalalgia*. 2012;32(2012):431-2.
- [15] Goadsby PJ. The vascular theory of migraine--a great story wrecked by the facts. *Brain*. 2009;132(2009):6-7.
- [16] Penfield W, McNaughton F. Dural headache and innervation of the dura mater. *Arch Neurol Psychiatry*. 1940;44(1940):43-75.
- [17] McNaughton FL, Feindel W. Innervation of intracranial structures: a reappraisal. In: Rose FC, editor *Physiological aspects of clinical neurology*. Oxford: Blackwell Scientific, 1977:279-93.
- [18] Millan MJ. Descending control of pain. *Prog Neurobiol*. 2002;66(2002):355-474.
- [19] Burstein R, Yamamura H, Malick A, Strassman AM. Chemical stimulation of the intracranial dura induces enhanced responses to facial stimulation in brain stem trigeminal neurons. *J-Neurophysiol*. 1998;79(1998):964-82.
- [20] Akerman S, Holland PR, Goadsby PJ. Cannabinoid (CB1) receptor activation inhibits trigeminovascular neurons. *J Pharmacol Exp Ther*. 2007;320(2007):64-71.

- [21] Liu Y, Broman J, Edvinsson L. Central projections of the sensory innervation of the rat middle meningeal artery. *Brain Res.* 2008;1208(2008):103-10.
- [22] Liu Y, Broman J, Edvinsson L. Central projections of sensory innervation of the rat superior sagittal sinus. *Neuroscience.* 2004;129(2004):431-7.
- [23] Holland PR, Akerman S, Goadsby PJ. Modulation of nociceptive dural input to the trigeminal nucleus caudalis via activation of the orexin 1 receptor in the rat. *Eur J Neurosci.* 2006;24(2006):2825-33.
- [24] Liu Y, Broman J, Zhang M, Edvinsson L. Brainstem and thalamic projections from a craniovascular sensory nervous centre in the rostral cervical spinal dorsal horn of rats. *Cephalalgia.* 2009;29(2009):935-48.
- [25] Matsushita M, Ikeda M, Okado N. The cells of origin of the trigeminothalamic, trigeminospinal and trigeminocerebellar projections in the cat. *Neuroscience.* 1982;7(1982):1439-54.
- [26] Shigenaga Y, Nakatani Z, Nishimori T, Suemune S, Kuroda R, Matano S. The cells of origin of cat trigeminothalamic projections: especially in the caudal medulla. *Brain Res.* 1983;277(1983):201-22.
- [27] Williams MN, Zahm DS, Jacquin MF. Differential foci and synaptic organization of the principal and spinal trigeminal projections to the thalamus in the rat. *Eur J Neurosci.* 1994;6(1994):429-53.
- [28] Veinante P, Jacquin MF, Deschenes M. Thalamic projections from the whisker-sensitive regions of the spinal trigeminal complex in the rat. *J Comp Neurol.* 2000;420(2000):233-43.

- [29] Robert C, Bourgeois L, Arreto CD, Condes-Lara M, Nosedá R, Jay T, Villanueva L. Paraventricular hypothalamic regulation of trigeminovascular mechanisms involved in headaches. *J Neurosci*. 2013;33(2013):8827-40.
- [30] Malick A, Burstein R. Cells of origin of the trigeminohypothalamic tract in the rat. *J Comp Neurol*. 1998;400(1998):125-44.
- [31] Malick A, Jakubowski M, Elmquist JK, Saper CB, Burstein R. A neurohistochemical blueprint for pain-induced loss of appetite. *Proc Natl Acad Sci U S A*. 2001;98(2001):9930-5.
- [32] Malick A, Strassman RM, Burstein R. Trigeminothalamic and reticulohypothalamic tract neurons in the upper cervical spinal cord and caudal medulla of the rat. *J Neurophysiol*. 2000;84(2000):2078-112.
- [33] Burstein R, Cliffer KD, Giesler GJ, Jr. Direct somatosensory projections from the spinal cord to the hypothalamus and telencephalon. *J Neurosci*. 1987;7(1987):4159-64.
- [34] Burstein R, Dado RJ, Giesler GJ, Jr. The cells of origin of the spinothalamic tract of the rat: a quantitative reexamination. *Brain Res*. 1990;511(1990):329-37.
- [35] Weiller C, May A, Limmroth V, Jüptner M, Kaube H, Schayck RV, et al. Brain stem activation in spontaneous human migraine attacks. *Nature medicine*. 1995;1(1995):658-60.
- [36] Afridi SK, Matharu MS, Lee L, Kaube H, Friston KJ, Frackowiak RS, Goadsby PJ. A PET study exploring the laterality of brainstem activation in migraine using glyceryl trinitrate. *Brain*. 2005;128(2005):932-9.
- [37] Maniyar FH, Sprenger T, Monteith T, Schankin C, Goadsby PJ. Brain activations in the premonitory phase of nitroglycerin-triggered migraine attacks. *Brain*. 2013(2013).
- [38] Schulte LH, May A. The migraine generator revisited: continuous scanning of the migraine cycle over 30 days and three spontaneous attacks. *Brain*. 2016;139(2016):1987-93.

- [39] Schulte LH, Sprenger C, May A. Physiological brainstem mechanisms of trigeminal nociception: An fMRI study at 3T. *Neuroimage*. 2016;124(2016):518-25.
- [40] Burstein R, Jakubowski M. Unitary hypothesis for multiple triggers of the pain and strain of migraine. *J Comp Neurol*. 2005;493(2005):9-14.
- [41] Edelmayer RM, Vanderah TW, Majuta L, Zhang ET, Fioravanti B, De Felice M, et al. Medullary pain facilitating neurons mediate allodynia in headache-related pain. *Annals of neurology*. 2009;65(2009):184-93.
- [42] Hoskin KL, Bulmer DCE, Lasalandra M, Jonkman A, Goadsby PJ. Fos expression in the midbrain periaqueductal grey after trigeminovascular stimulation. *J Anat*. 2001;198(2001):29-35.
- [43] Knight YE, Classey JD, Lasalandra MP, Akerman S, Kowacs F, Hoskin KL, Goadsby PJ. Patterns of fos expression in the rostral medulla and caudal pons evoked by noxious craniovascular stimulation and periaqueductal gray stimulation in the cat. *Brain Res*. 2005;1045(2005):1-11.
- [44] Lambert GA, Hoskin KL, Zagami AS. Cortico-NRM influences on trigeminal neuronal sensation. *Cephalalgia*. 2008;28(2008):640-52.
- [45] Holland PR. Biology of Neuropeptides: Orexinergic Involvement in Primary Headache Disorders. *Headache*. 2017;57 Suppl 2(2017):76-88.
- [46] Schneeberger M, Gomis R, Claret M. Hypothalamic and brainstem neuronal circuits controlling homeostatic energy balance. *J Endocrinol*. 2014;220(2014):T25-46.
- [47] Shinpo K, Hirai Y, Maezawa H, Totsuka Y, Funahashi M. The role of area postrema neurons expressing H-channels in the induction mechanism of nausea and vomiting. *Physiol Behav*. 2012;107(2012):98-103.



- [48] Lutz TA, Senn M, Althaus J, Del Prete E, Ehrensperger F, Scharrer E. Lesion of the area postrema/nucleus of the solitary tract (AP/NTS) attenuates the anorectic effects of amylin and calcitonin gene-related peptide (CGRP) in rats. *Peptides*. 1998;19(1998):309-17.
- [49] Price CJ, Hoyda TD, Ferguson AV. The area postrema: a brain monitor and integrator of systemic autonomic state. *Neuroscientist*. 2008;14(2008):182-94.
- [50] Giffin NJ, Lipton RB, Silberstein SD, Olesen J, Goadsby PJ. The migraine postdrome: An electronic diary study. *Neurology*. 2016;87(2016):309-13.
- [51] Holland PR. Headache and sleep: shared pathophysiological mechanisms. *Cephalalgia*. 2014;34(2014):725-44.
- [52] Bower RL, Eftekhari S, Waldvogel HJ, Faull RL, Tajti J, Edvinsson L, et al. Mapping the calcitonin receptor in human brain stem. *Am J Physiol Regul Integr Comp Physiol*. 2016;310(2016):R788-93.
- [53] Eftekhari S, Gaspar RC, Roberts R, Chen TB, Zeng Z, Villarreal S, et al. Localization of CGRP receptor components and receptor binding sites in rhesus monkey brainstem: A detailed study using in situ hybridization, immunofluorescence, and autoradiography. *J Comp Neurol*. 2016;524(2016):90-118.
- [54] Ma W, Chabot JG, Powell KJ, Jhamandas K, Dickerson IM, Quirion R. Localization and modulation of calcitonin gene-related peptide-receptor component protein-immunoreactive cells in the rat central and peripheral nervous systems. *Neuroscience*. 2003;120(2003):677-94.
- [55] Tajti J, Uddman R, Edvinsson L. Neuropeptide localization in the "migraine generator" region of the human brainstem. *Cephalalgia*. 2001;21(2001):96-101.

- [56] Akerman S, Holland PR, Lasalandra MP, Goadsby PJ. Endocannabinoids in the brainstem modulate dural trigeminovascular nociceptive traffic via CB1 and "triptan" receptors: implications in migraine. *J Neurosci*. 2013;33(2013):14869-77.
- [57] Amara SG, Arriza JL, Leff SE, Swanson LW, Evans RM, Rosenfeld MG. Expression in brain of a messenger RNA encoding a novel neuropeptide homologous to calcitonin gene-related peptide. *Science*. 1985;229(1985):1094-7.
- [58] Bartsch T, Knight YE, Goadsby PJ. Activation of 5-HT(1B/1D) receptor in the periaqueductal gray inhibits nociception. *Annals of neurology*. 2004;56(2004):371-81.
- [59] Goadsby PJ, Duckworth JW. Low frequency stimulation of the locus coeruleus reduces regional cerebral blood flow in the spinalized cat. *Brain Res*. 1989;476(1989):71-7.
- [60] Kelman L. The triggers or precipitants of the acute migraine attack. *Cephalalgia*. 2007;27(2007):394-402.
- [61] de Souza E, Covenas R, Yi P, Aguilar LA, Lerma L, Andrade R, et al. Mapping of CGRP in the alpaca (*Lama pacos*) brainstem. *J Chem Neuroanat*. 2008;35(2008):346-55.
- [62] Warfvinge K, Edvinsson L. Distribution of CGRP and CGRP receptor components in the rat brain. *Cephalalgia*. 2017(2017):333102417728873.
- [63] Inagaki S, Kito S, Kubota Y, Girgis S, Hillyard CJ, MacIntyre I. Autoradiographic localization of calcitonin gene-related peptide binding sites in human and rat brains. *Brain Res*. 1986;374(1986):287-98.
- [64] Pozo-Rosich P, Storer RJ, Charbit AR, Goadsby PJ. Periaqueductal gray calcitonin gene-related peptide modulates trigeminovascular neurons. *Cephalalgia*. 2015;35(2015):1298-307.
- [65] Bhatt DK, Gupta S, Ploug KB, Jansen-Olesen I, Olesen J. mRNA distribution of CGRP and its receptor components in the trigeminovascular system and other pain related structures in

rat brain, and effect of intracerebroventricular administration of CGRP on Fos expression in the TNC. *Neurosci Lett*. 2014;559(2014):99-104.

[66] Van Rossum D, Menard DP, Fournier A, St-Pierre S, Quirion R. Binding profile of a selective calcitonin gene-related peptide (CGRP) receptor antagonist ligand, [125I-Tyr]hCGRP8-37, in rat brain and peripheral tissues. *J Pharmacol Exp Ther*. 1994;269(1994):846-53.

[67] Nemoto T, Konno A, Chiba T. Synaptic contact of neuropeptide-and amine-containing axons on parasympathetic preganglionic neurons in the superior salivatory nucleus of the rat. *Brain Res*. 1995;685(1995):33-45.

[68] Xiaocheng W, Zhaohui S, Junhui X, Lei Z, Lining F, Zuoming Z. Expression of calcitonin gene-related peptide in efferent vestibular system and vestibular nucleus in rats with motion sickness. *PLoS One*. 2012;7(2012):e47308.

[69] Ahn SK, Khalmuratova R, Jeon SY, Kim JP, Park JJ, Hur DG, Balaban CD. Colocalization of 5-HT<sub>1F</sub> receptor and calcitonin gene-related peptide in rat vestibular nuclei. *Neurosci Lett*. 2009;465(2009):151-6.

[70] Mulderry PK, Ghatei MA, Spokes RA, Jones PM, Pierson AM, Hamid QA, et al. Differential expression of alpha-CGRP and beta-CGRP by primary sensory neurons and enteric autonomic neurons of the rat. *Neuroscience*. 1988;25(1988):195-205.

[71] Harmann PA, Chung K, Briner RP, Westlund KN, Carlton SM. Calcitonin gene-related peptide (CGRP) in the human spinal cord: a light and electron microscopic analysis. *J Comp Neurol*. 1988;269(1988):371-80.

[72] van Rossum D, Hanisch UK, Quirion R. Neuroanatomical localization, pharmacological characterization and functions of CGRP, related peptides and their receptors. *Neuroscience and biobehavioral reviews*. 1997;21(1997):649-78.

- [73] Edvinsson L. The Trigeminovascular Pathway: Role of CGRP and CGRP Receptors in Migraine. *Headache*. 2017;57 Suppl 2(2017):47-55.
- [74] Hay DL, Poyner DR, Smith DM. Desensitisation of adrenomedullin and CGRP receptors. *Regul Pept*. 2003;112(2003):139-45.
- [75] Walker CS, Eftekhari S, Bower RL, Wilderman A, Insel PA, Edvinsson L, et al. A second trigeminal CGRP receptor: function and expression of the AMY1 receptor. *Ann Clin Transl Neurol*. 2015;2(2015):595-608.
- [76] Hagner S, Haberberger RV, Overkamp D, Hoffmann R, Voigt KH, McGregor GP. Expression and distribution of calcitonin receptor-like receptor in human hairy skin. *Peptides*. 2002;23(2002):109-16.
- [77] Mallee JJ, Salvatore CA, LeBourdelles B, Oliver KR, Longmore J, Koblan KS, Kane SA. Receptor activity-modifying protein 1 determines the species selectivity of non-peptide CGRP receptor antagonists. *J Biol Chem*. 2002;277(2002):14294-8.
- [78] Luebke AE, Dahl GP, Roos BA, Dickerson IM. Identification of a protein that confers calcitonin gene-related peptide responsiveness to oocytes by using a cystic fibrosis transmembrane conductance regulator assay. *Proc Natl Acad Sci U S A*. 1996;93(1996):3455-60.
- [79] Olesen J, Diener HC, Husstedt IW, Goadsby PJ, Hall D, Meier U, et al. Calcitonin gene-related peptide receptor antagonist BIBN 4096 BS for the acute treatment of migraine. *N Engl J Med*. 2004;350(2004):1104-10.
- [80] Tfelt-Hansen P, Olesen J. Possible site of action of CGRP antagonists in migraine. *Cephalalgia*. 2011;31(2011):748-50.
- [81] Sixt ML, Messlinger K, Fischer MJ. Calcitonin gene-related peptide receptor antagonist olcegepant acts in the spinal trigeminal nucleus. *Brain*. 2009;132(2009):3134-41.

- [82] Hostetler ED, Joshi AD, Sanabria-Bohorquez S, Fan H, Zeng ZZ, Purcell M, et al. In Vivo Quantification of Calcitonin Gene-Related Peptide Receptor Occupancy by Telcagepant in Rhesus Monkey and Human Brain Using the Positron Emission Tomography Tracer [C-11]MK-4232. *J Pharmacol Exp Ther.* 2013;347(2013):478-86.
- [83] Goadsby PJ, Reuter U, Hallstrom Y, Broessner G, Bonner JH, Zhang F, et al. A Controlled Trial of Erenumab for Episodic Migraine. *N Engl J Med.* 2017;377(2017):2123-32.
- [84] Schankin CJ, Maniyar FH, Seo Y, Kori S, Eller M, Chou DE, et al. Ictal lack of binding to brain parenchyma suggests integrity of the blood-brain barrier for <sup>11</sup>C-dihydroergotamine during glyceryl trinitrate-induced migraine. *Brain.* 2016;139(2016):1994-2001.
- [85] Benarroch EE. Circumventricular organs: receptive and homeostatic functions and clinical implications. *Neurology.* 2011;77(2011):1198-204.
- [86] Farkkila M, Diener HC, Geraud G, Lainez M, Schoenen J, Harner N, et al. Efficacy and tolerability of lasmiditan, an oral 5-HT<sub>1F</sub> receptor agonist, for the acute treatment of migraine: a phase 2 randomised, placebo-controlled, parallel-group, dose-ranging study. *Lancet Neurol.* 2012;11(2012):405-13.
- [87] Iyengar S, Ossipov MH, Johnson KW. The role of CGRP in peripheral and central pain mechanisms including migraine. *Pain.* 2016(2016).
- [88] Eftekhari S, Salvatore CA, Calamari A, Kane SA, Tajti J, Edvinsson L. Differential distribution of calcitonin gene-related peptide and its receptor components in the human trigeminal ganglion. *Neuroscience.* 2010;169(2010):683-96.
- [89] Tiller-Borich JK, Capili H, Gordan GS. Human brain calcitonin gene-related peptide (CGRP) is concentrated in the locus caeruleus. *Neuropeptides.* 1988;11(1988):55-61.

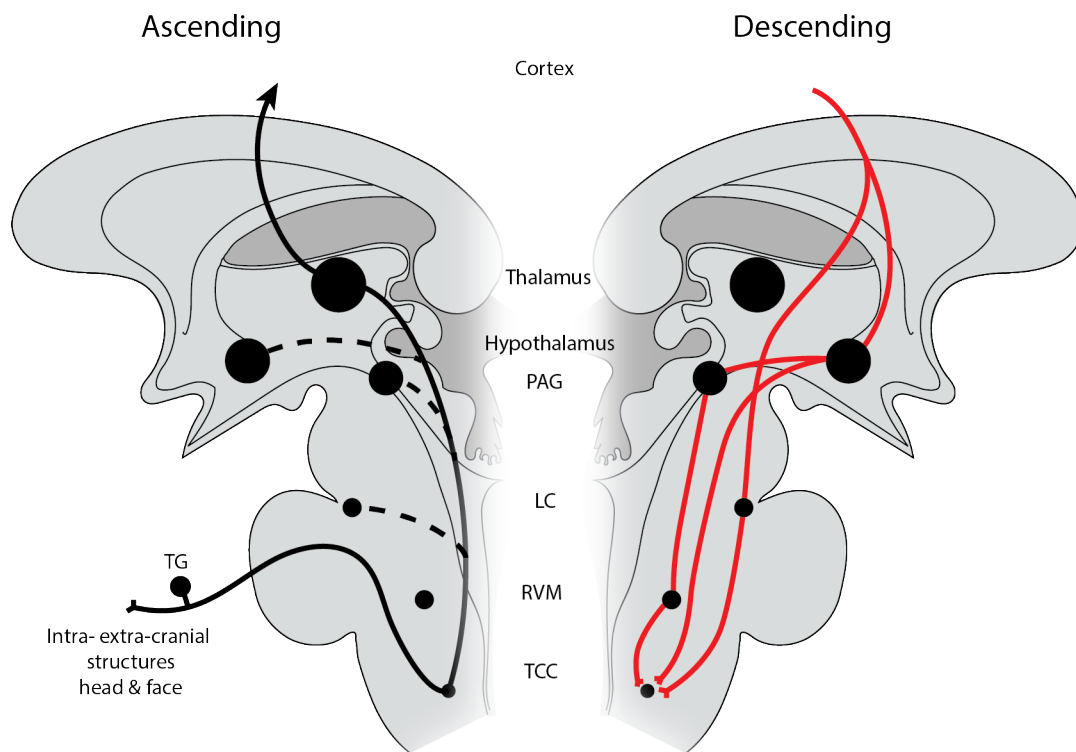
- [90] Storer RJ, Akerman S, Goadsby PJ. Calcitonin gene-related peptide (CGRP) modulates nociceptive trigeminovascular transmission in the cat. *Br J Pharmacol.* 2004;142(2004):1171-81.
- [91] Kagenneck C, Nixdorf-Bergweiler BE, Messlinger K, Fischer MJ. Release of CGRP from mouse brainstem slices indicates central inhibitory effect of triptans and kynurenate. *J Headache Pain.* 2014;15(2014):7.
- [92] Carter ME, Yizhar O, Chikahisa S, Nguyen H, Adamantidis A, Nishino S, et al. Tuning arousal with optogenetic modulation of locus coeruleus neurons. *Nat Neurosci.* 2010;13(2010):1526-33.
- [93] Samuels ER, Szabadi E. Functional neuroanatomy of the noradrenergic locus coeruleus: its roles in the regulation of arousal and autonomic function part II: physiological and pharmacological manipulations and pathological alterations of locus coeruleus activity in humans. *Curr Neuropharmacol.* 2008;6(2008):254-85.
- [94] McCall JG, Al-Hasani R, Siuda ER, Hong DY, Norris AJ, Ford CP, Bruchas MR. CRH Engagement of the Locus Coeruleus Noradrenergic System Mediates Stress-Induced Anxiety. *Neuron.* 2015;87(2015):605-20.
- [95] Llorca-Torralba M, Borges G, Neto F, Mico JA, Berrocoso E. Noradrenergic Locus Coeruleus pathways in pain modulation. *Neuroscience.* 2016;338(2016):93-113.
- [96] Goadsby PJ, Lambert GA, Lance JW. Differential effects on the internal and external carotid circulation of the monkey evoked by locus coeruleus stimulation. *Brain Res.* 1982;249(1982):247-54.
- [97] Goadsby PJ, Lambert GA, Lance JW. Effects of locus coeruleus stimulation on carotid vascular resistance in the cat. *Brain Res.* 1983;278(1983):175-83.

- [98] Boyer N, Signoret-Genest J, Artola A, Dallel R, Monconduit L. Propranolol treatment prevents chronic central sensitization induced by repeated dural stimulation. *Pain*. 2017(2017).
- [99] Tassorelli C, Joseph SA. Systemic nitroglycerin induces Fos immunoreactivity in brainstem and forebrain structures of the rat. *Brain Res*. 1995;682(1995):167-81.
- [100] Sawada H, Yamaguchi H, Shimbara T, Toshinai K, Mondal MS, Date Y, et al. Central effects of calcitonin receptor-stimulating peptide-1 on energy homeostasis in rats. *Endocrinology*. 2006;147(2006):2043-50.
- [101] Lane S, Roloff E, Fernandes F, Wheildon G, Brennan L, Thompson G, et al. Olcegepant potentiates astrocyte-mediated noradrenaline release in the locus coeruleus. *FENS*. Milan, Italy, 2014.
- [102] Aston-Jones G, Bloom FE. Activity of norepinephrine-containing locus coeruleus neurons in behaving rats anticipates fluctuations in the sleep-waking cycle. *J Neurosci*. 1981;1(1981):876-86.
- [103] Vila-Pueyo M, Strother L, Goadsby PJ, Holland PR. The Role of the Locus Coeruleus in Regulating Trigemino-vascular Nociception. *Cephalalgia*. 2016;36(2016):152-.
- [104] Jia Z, Tang W, Zhao D, Yu S. Disrupted functional connectivity between the periaqueductal gray and other brain regions in a rat model of recurrent headache. *Sci Rep*. 2017;7(2017):3960.
- [105] Knight YE, Bartsch T, Goadsby PJ. Trigeminal antinociception induced by bicuculline in the periaqueductal gray (PAG) is not affected by PAG P/Q-type calcium channel blockade in rat. *Neurosci Lett*. 2003;336(2003):113-6.

- [106] Knight YE, Bartsch T, Kaube H, Goadsby PJ. P/Q-type calcium-channel blockade in the periaqueductal gray facilitates trigeminal nociception: a functional genetic link for migraine? *J Neurosci.* 2002;22(2002):RC213.
- [107] Knight YE, Goadsby PJ. The periaqueductal grey matter modulates trigeminovascular input: a role in migraine? *Neuroscience.* 2001;106(2001):793-800.
- [108] Cumberbatch MJ, Hill RG, Hargreaves RJ. Rizatriptan has central antinociceptive effects against durally evoked responses. *Eur J Pharmacol.* 1997;328(1997):37-40.
- [109] Yao G, Han X, Hao T, Huang Q, Yu T. Effects of rizatriptan on the expression of calcitonin gene-related peptide and cholecystokinin in the periaqueductal gray of a rat migraine model. *Neurosci Lett.* 2015;587(2015):29-34.
- [110] Huang Y, Brodda-Jansen G, Lundeberg T, Yu LC. Anti-nociceptive effects of calcitonin gene-related peptide in nucleus raphe magnus of rats: an effect attenuated by naloxone. *Brain Res.* 2000;873(2000):54-9.
- [111] Basbaum AI, Clanton CH, Fields HL. Three bulbospinal pathways from the rostral medulla of the cat: an autoradiographic study of pain modulating systems. *J Comp Neurol.* 1978;178(1978):209-24.
- [112] Holstege G, Kuypers HG. The anatomy of brain stem pathways to the spinal cord in cat. A labeled amino acid tracing study. *Prog Brain Res.* 1982;57(1982):145-75.
- [113] Fields HL, Heinricher MM. Anatomy and physiology of a nociceptive modulatory system. *Philos Trans R Soc Lond B Biol Sci.* 1985;308(1985):361-74.
- [114] Foo H, Mason P. Brainstem modulation of pain during sleep and waking. *Sleep Med Rev.* 2003;7(2003):145-54.

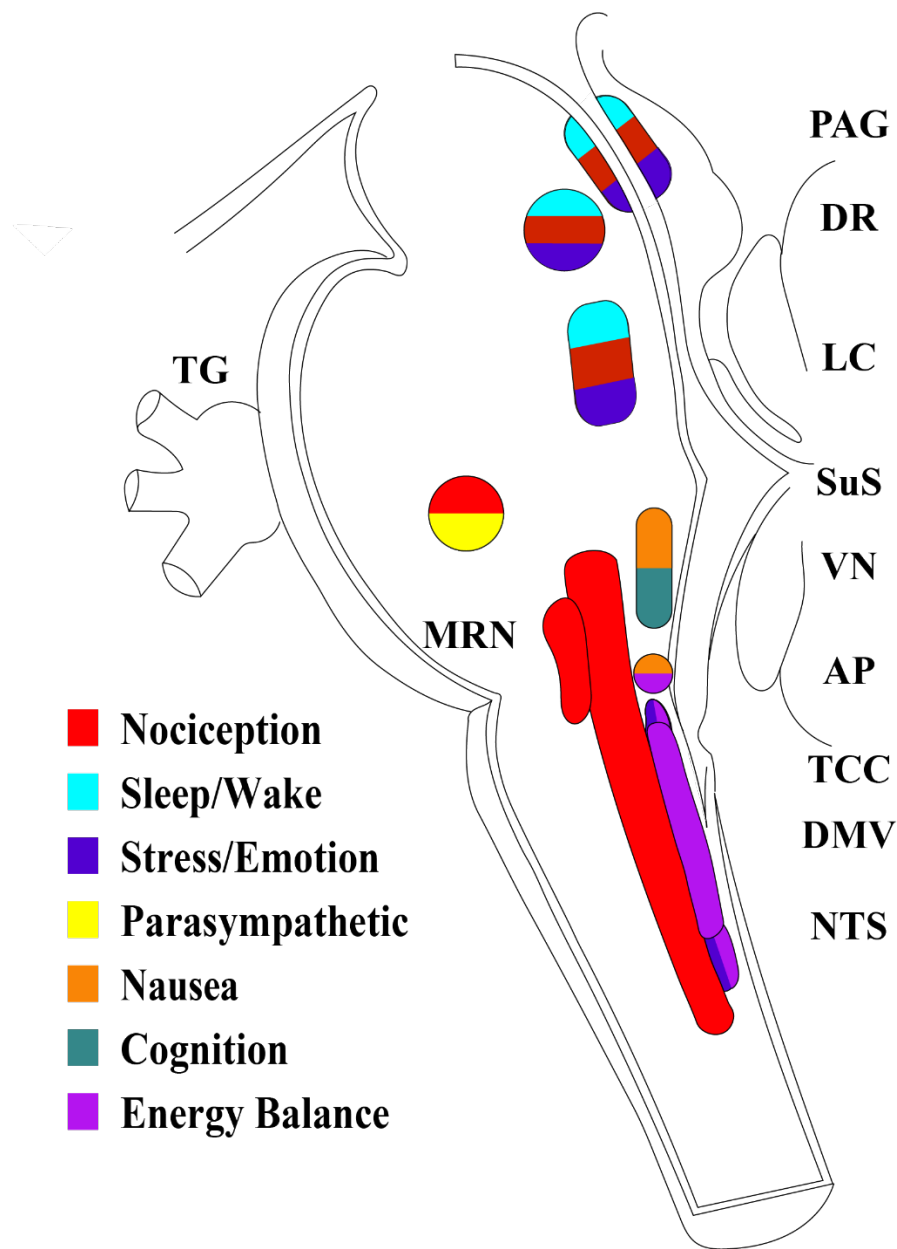


- [115] Foo H, Mason P. Sensory suppression during feeding. *Proc Natl Acad Sci U S A*. 2005;102(2005):16865-9.
- [116] Roeder Z, Chen Q, Davis S, Carlson JD, Tupone D, Heinricher MM. Parabrachial complex links pain transmission to descending pain modulation. *Pain*. 2016;157(2016):2697-708.
- [117] Stolte B, Holle D, Naegel S, Diener HC, Obermann M. Vestibular migraine. *Cephalalgia*. 2015;35(2015):262-70.
- [118] Kong WJ, Scholtz AW, Hussl B, Kammen-Jolly K, Schrott-Fischer A. Localization of efferent neurotransmitters in the inner ear of the homozygous Bronx waltzer mutant mouse. *Hear Res*. 2002;167(2002):136-55.
- [119] Kong WJ, Scholtz AW, Kammen-Jolly K, Gluckert R, Hussl B, von Cauvenberg PB, Schrott-Fischer A. Ultrastructural evaluation of calcitonin gene-related peptide immunoreactivity in the human cochlea and vestibular endorgans. *Eur J Neurosci*. 2002;15(2002):487-97.
- [120] Wackym PA, Popper P, Ward PH, Micevych PE. Cell and molecular anatomy of nicotinic acetylcholine receptor subunits and calcitonin gene-related peptide in the rat vestibular system. *Otolaryngol Head Neck Surg*. 1991;105(1991):493-510.
- [121] Wang XC, Shi ZH, Xue JH, Zhang L, Feng LN, Zhang ZM. Expression of Calcitonin Gene-Related Peptide in Efferent Vestibular System and Vestibular Nucleus in Rats with Motion Sickness. *Plos One*. 2012;7(2012).
- [122] Luebke AE, Holt JC, Jordan PM, Wong YS, Caldwell JS, Cullen KE. Loss of alpha-calcitonin gene-related peptide (alphaCGRP) reduces the efficacy of the Vestibulo-ocular Reflex (VOR). *J Neurosci*. 2014;34(2014):10453-8.
- [123] Wang N, Huang HL, Zhou H, Yu CY. Cognitive impairment and quality of life in patients with migraine-associated vertigo. *Eur Rev Med Pharmacol Sci*. 2016;20(2016):4913-7.



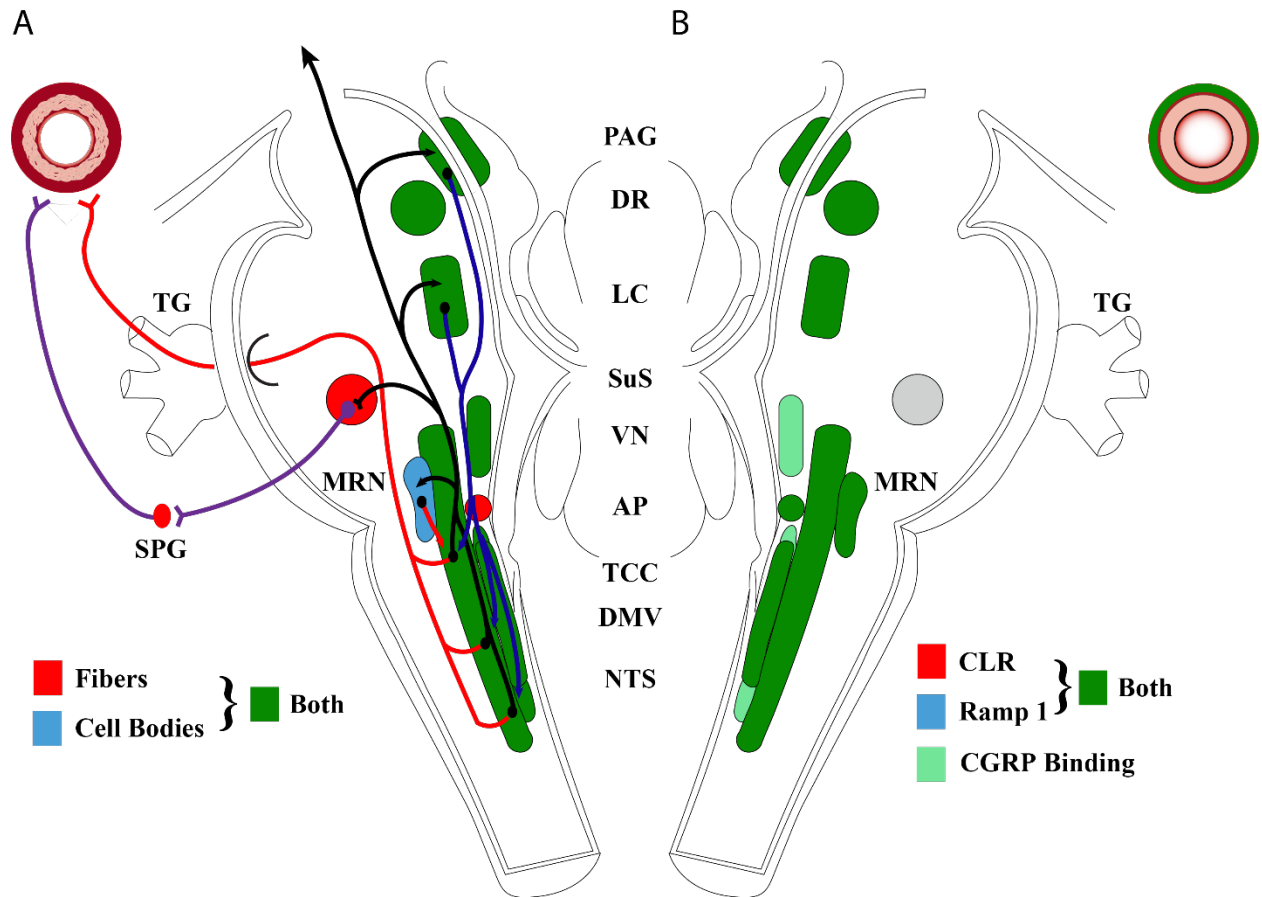
**Figure 1: Ascending and descending projections via the brainstem in migraine.**

The trigeminal ganglion (TG) gives rise to the pseudo-unipolar trigeminal primary afferents which synapse on the intra- and extra-cranial structures (blood vessels and dura mater) as well as the spinal cord trigeminocervical complex (TCC). Second-order neurons from the TCC ascend in the trigeminothalamic tract synapsing on third-order thalamocortical neurons before passing to the cortex. Direct and indirect ascending projections additionally exist to key brainstem and diencephalic nuclei including the locus coeruleus (LC), periaqueductal grey (PAG) and hypothalamus. The TCC is under the regulation of direct and indirect descending modulatory pathways. Direct and indirect projections (via the hypothalamus) exist from the cortex, whilst key brainstem nuclei including the PAG, LC and rostral ventromedial medulla (RVM) provide modulatory pathways that arise within the nuclei or are under top down regulation. This complex network of descending modulatory circuits potentially regulates the TCC providing pro- and anti-nociceptive drive.



**Figure 2: The functional importance of selected brainstem nuclei with respect to migraine-associated symptoms.**

AP; area postrema, DMV; dorsal motor nucleus of the vagal, DR; dorsal raphe, LC; locus coeruleus, MRN, nucleus raphe magnus, NTS; nucleus of the solitary tract, PAG; periaqueductal gray, SPG; sphenopalatine ganglion, SuS; superior salivatory nucleus, TCC; trigeminocervical complex, TG; trigeminal ganglion, VN; vestibular nuclei.



**Figure 3: Overview of CGRP expression in selected brainstem nuclei.**

(A) The expression of CGRP fibers (red), cell bodies (soma, blue) or both (green) in selected brainstem nuclei are shown. (B) The expression of calcitonin receptor-like receptor (CLR, red), receptor activity modifying protein 1 (RAMP 1, blue) or both (green) in selected brainstem nuclei are shown. Where CLR and RAMP 1 expression is unknown, CGRP receptor binding has been used to highlight the presence of functional CGRP receptors (light green). The above information is taken from the references and data presented in Table 1. AP; area postrema, DMV; dorsal motor nucleus of the vagal, DR; dorsal raphe, LC; locus coeruleus, MRN, nucleus raphe magnus, NTS; nucleus of the solitary tract, PAG; periaqueductal gray, SPG; sphenopalatine ganglion, SuS; superior salivatory nucleus, TCC; trigeminocervical complex, TG; trigeminal ganglion, VN; vestibular nuclei.